1-(N-Trifluoroacetylamino)alkylphosphonic acids: synthesis and properties

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Summary. The 1-(N-trifluoroacetylamino)alkylphosphonic acids (TFA-AA^P) – sub-products in the synthesis of O,O-dialkyl 1-(N-trifluoroacetylamino)alkylphosphonates and O,O-diethyl 1-aminoalkylphosphonates, were synthesized in two-stage transformations of 1-aminoalkylphosphonic acids including: trifluoroacetylation of 1-aminoalkylphosphonic acids (AAP) using a trifluoroacetic anhydride/trifluoroacetic acid reagent $(AA^P + TFAA/TFA \rightarrow 2)$ and subsequent hydrolysis of the intermediary compounds 2 into desired TFA-AAP (2-TFA-AAP). These intermediates 2 presented mixtures of the type of mixed anhydrides of TFAA and 1-(Ntrifluoroacetylamino)alkylphosphonic, pyrophosphonic and polyphosphonic acids, which underwent rapid and quantitative conversion to corresponding TFA-AAP during treatment with an excess of water. The title acids were isolated by direct evaporation of the corresponding postreaction mixtures, and their physicochemical proprieties, including deacylation abilities, were determined. TFA-AAP compounds can be reconverted into the starting amino acids AAP under respectively mild conditions $(AA^P \rightarrow TFA - AA^P \rightarrow AA^P)$.

Keywords: Acylation – Trifluoroacetylation – Mixed anhydrides – Aminophosphonic acids – 1-Aminoalkylphosphonic acids – 1-(*N*-Trifluoroacetylamino)alkylphosphonic acids

Introduction

The 1-(*N*-trifluoroacetylamino)alkylphosphonic acids (TFA-AA^P) belong to an interesting (Kafarski and Mastalerz, 1984; Kafarski and Lejczak, 1991; Kukhar and Hudson, 2000), although almost not explored group of *N*-acyl derivatives of aminophosphonic acids (AA^P), as until now only TFA-Val^P was synthesized and partly characterized (Khomutov et al., 1979).

These 1-(*N*-trifluoroacetylamino)alkylphosphonic acids are sub-products in the synthesis of *O*,*O*-dialkyl 1-(*N*-trifluoroacetylamino)alkylphosphonates (Kudzin and Łuczak, 1995) and *O*,*O*-diethyl 1-aminoalkylphosphonates (Kudzin et al., 1997), important key-substrates in the preparation of

phosphonopeptides (Kafarski et al., 1984). 1-(*N*-Trifluoro-acetylamino)alkylphosphonic acids, due to liability of the TFA-NH linkage are also potentially attractive substrates in the synthesis of optically active 1-aminoalkylphosphonic acids (Dahvan and Redmore, 1987; Kukhar and Hudson, 2000).

In this communication we present a simple and quantitative method of conversion of 1-aminoalkylphosphonic acids into the title 1-(N-trifluoroacetylamino)alkylphosphonic acids $(AA^P \rightarrow 2 \rightarrow TFA-AA^P)$ and tentative results on the course of trifluoroacetylation $(AA^P \rightarrow 2)$. In addition, also results on conditions of the re-conversion of 1-(N-trifluoroacetylamino)-alkylphosphonic acids into the starting 1-aminoalkylphosphonic acids, i.e. on deprotections of the amine function in TFA-N-protected amino acids $(AA^P \rightarrow TFA-AA^P \rightarrow AA^P)$.

Materials and methods

Materials

1-Aminoalkylphosphonic acids were prepared according to Kudzin and Stec [1978 (Ala^P, Val^P, Pgly^P and Phe^P)] and Soroka [1989 (Gly^P)]. All other reagents were purchased from Aldrich (Milwaukee, IL, USA). Acetate buffer solution contained 10 mmol AcOH and 10 mmol of AcONa in 5 ml of aqueous solution.

Fig. 1. Structure of 1-(N-trifluoroacetylamino)alkylphosphonic acids TFA-AA P (3)

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Z. H. Kudzin et al.

Equipment

All melting points were determined using a MEL-TEMP II capillary melting point apparatus (Laboratory Devices Inc., USA) and are uncorrected. NMR spectra were recorded on a Brucker AC 200 spectrometer operating at 200 MHz for ^{1}H NMR, at 188.3 MHz for ^{19}F NMR and at 81.01 MHz for ^{31}P NMR. Elemental analyses (C, H) were recorded on an Elemental Analyzer Euro EA (Eurovector, Italy). The chemical shift data for each signal on ^{1}H NMR are taken in units of δ relative to CHCl₃ (δ 7.26) for TFA-CDCl₃ solutions and H₂O (δ 4.72) for aqueous solutions, The chemical shifts of ^{31}P are recorded relative to external 85% H₃PO₄ (δ 0.0) with broad-band ^{1}H decoupling.

General procedure of N-trifluoroacetylation of 1-aminoalkylphosphonic acids

A sample of AA^P (1 mmol) was added to a solution of TFA (0.5 ml) and TFAA (1 ml, 1.51 g; 7.2 mmol), and the mixture was stirred at ambient temperature until homogenization, and then for additional 1 h. The formed solution was evaporated (20 °C, 10–20 mmHg), an oily residue was dissolved in cold water (5 ml), and evaporated again *to dryness* (50 °C, 10–20 mmHg). The residues presented pure 1-(*N*-trifluoroacetylamino)-alkylphosphonic acids, which were stored in a vacuum desiccator over solid KOH.

TFA-Gly^P (3a)

Yield: 96–98% (100% according to ^{31}P NMR). Colorless, hygroscopic crystals; mp 66–68 °C. ^{31}P NMR (δ_B ppm): 18.1 (2 M HCl/D₂O), 16.8 (D₂O), 19.3 (DMF/CDCl₃). ^{19}F NMR (δ_F ppm): -74.88 (D₂O). ^{1}H NMR (δ_H , ppm): δ_H (CDCl₃): 3.71 (1H, dd, J 5.6, 13.1 Hz, TFANHCH₂P(O)(OH₂)], 7.91–8.07 (1H, m, TFANHCH₂P), 11.0–11.31 [2H, br s, CH₂P(O)(OH₂)]; δ_H [CDCl₃-TFA (3:2)]: 4.05 [2H, d, J 6.1 Hz, TFANHCH₂P(O)(OH₂)], 7.00–7.50 (1H, br s, TFANHCH₂P). Found: C 17.37, H 2.57, P 14.65. Calc. for C₃H₃F₃NPO₄ [207.1]: C 17.40, H 2.42, P 14.96.

TFA-Ala^P (3b)

Yield: 96–98% (100% according to ^{31}P NMR). Colorless, hygroscopic crystals; mp 102–104°C. ^{31}P NMR ($\delta_{\rm P}$ ppm): 21.2 (2 M HCl/D₂O), 20.3 (D₂O), 19.3 (DMF/CDCl₃). ^{19}F NMR ($\delta_{\rm F}$ ppm): $\delta_{\rm F}$ –74.81 (D₂O). ^{1}H NMR ($\delta_{\rm H}$ ppm): $\delta_{\rm H}$ (CDCl₃): 1.43 [3H, dd, J 7.2, 16.2 Hz, TFANHCH(C<u>H</u>₃)P(O)(OH)₂], 4.25–4.50 (1H, m, CH₃C<u>H</u>P), 7.56 (1H, d, J 8.3 Hz, TFANHCH), 8.25–8.50 [2H, br s, CHP(O)(O<u>H</u>₂)]; $\delta_{\rm H}$ [CDCl₃-TFA (3:2)]: 1.60 [3H, d, J 13.6 Hz, TFANHCH(C<u>H</u>₃)P(O)(OH)₂], 2.10–2.60 (1H, m, CH₃CHP), 7.55–7.85 (1H, m, TFAN<u>H</u>CHP). Found: C 21.50, H 3.40, P 14.30. Calc. for C₄H₇F₃NPO₄ [221.1]: C 21.73, H 3.19, P 14.01.

TFA-Val^P (3c)

Yield: 96–98% (100% according to ^{31}P NMR). Colorless, hygroscopic crystals; mp 164–166 °C [235–238 °C (Khomutov et al., 1979)]. ^{31}P NMR (δ_P ppm): 19.8 (2 M HCl/D₂O), 18.9 (D₂O), 18.2 (DMF/CDCl₃); ^{19}F NMR (δ_F ppm): -74.40 (D₂O). ^{1}H NMR (δ_H, ppm): δ_{H} [CDCl₃-TFA (3:2)]: 1.12 {6H, dd, J 7.0, 8.7 Hz, TFANH[(CH₃)₂CH]CHP(O)(OH)₂}, 2.12–2.40 [1H, m, (CH₃)₂CHCHP], 4.24–4.56 [1H, m, (CH₃)₂CHCHP], 7.65 (1H, d, J 7.0 Hz, TFANHCHP). Found: C 28.90, H 4.43, P 12.07. Calc. for C₆H₁₁F₃NPO₄ [249.1]: C 28.93, H 4.45, P 12.43.

TFA-Pgly^P (3d)

Yield: 96–98% (100% according to 31 P NMR). Colorless, hygroscopic crystals; mp 184–186 °C. 31 P NMR ($\delta_{\rm P}$ ppm): 16.1 (2 M HCl/D₂O), 14.8 (D₂O), 15.0 (DMF/CDCl₃). 19 F NMR ($\delta_{\rm F}$ ppm): -74.56 (D₂O). 1 H NMR

 $\begin{array}{l} (\delta_{H},\,ppm)\colon \delta_{H}\;(CDCl_{3});\; 5.36\;[1H,\,dd,\,J\;9.0,\;21.2\,Hz,\;TFANHC\underline{H}(C_{6}H_{5})-P(O)(OH)_{2}],\; 7.22-7.50\;[(5H,\,m)\colon 7.22-7.32\;(3H_{ar},\,m);\; 7.32-7.50\;(2H_{ar},\,m)],\; 8.26\;(1H,\,dd,\,J\;5.0,\;8.6\,Hz,\;TFA-N\underline{H}CH),\; 11.86-12.16\;[2H,\,br\;s,\;CHP(O)(O\underline{H}_{2})];\;\;\delta_{H}\;\;[CDCl_{3}\text{-}TFA\;(3:2)]\colon 5.66\;[1H,\,dd,\,J\;7.9,\;20.5\,Hz,\;TFANHC\underline{H}(C_{6}H_{5})P(O)(OH)_{2}],\; 7.32-7.50\;(5H_{ar},\,m),\; 8.0-8.25\;(1H,\,br\;s,\;TFAN\underline{H}CHP).\;Found:\;C\;38.58,\;H\;3.46,\;P\;11.15.\;Calc.\;for\;C_{9}H_{9}F_{3}NPO_{4}\;[283.1];\;C\;38.18,\;H\;3.20,\;P\;10.94. \end{array}$

TFA-PheP (3e)

Yield: 96–98% (100% according to ^{31}P NMR). Colorless, hygroscopic crystals; mp 146–150°C. ^{31}P NMR ($\delta_{\rm P}$ ppm): 18.9 (2 M HCl/D₂O), 17.9 (D₂O), 17.8 (DMF/CDCl₃). ^{19}F NMR ($\delta_{\rm F}$ ppm): -74.69 (D₂O) ^{1}H NMR ($\delta_{\rm H}$, ppm): $\delta_{\rm H}$ (CDCl₃): 2.91–5.08 [1H, m, TFA-NH(C₆H₅C<u>H</u>₂)-CHP(O)(OH)₂], 3.20–3.37 (1H, m, C₆H₅C<u>H</u>₂CHP), 4.46–4.65 (1H, m, PhCH₂C<u>H</u>P); 7.08–7.25 (m, 5H_{ar}), 8.04 (1H, d, J 9.7 Hz, TFAN<u>H</u>CH), 11.40–11.70 [2H, br s, CHP(O)(O<u>H</u>₂)]; $\delta_{\rm H}$ [CDCl₃-TFA (3:2)]: 2.90–3.10 [1H, m, TFANH(C₆H₅C<u>H</u>₂)CH-P(O)(OH)₂], 3.25–3.45 (1H, m, C₆H₅C<u>H</u>₂CHP), 4.60–4.90 (1H, m, PhCH₂C<u>H</u>P), 7.11–7.45 [(5H): 7.11–7.25 (3H_{ar}, m); 7.30–7.45 (2H_{ar}, m)], 7.54–7.90 (1H, br s, TFAN<u>H</u>CHP); Found: C 40.24, H 3.95, P 10.43. Calc. for C₁₀H₁₁F₃NPO₄ [297.2]: C 40.23, H 3.72, P 10.42.

Deacylation investigations of 1-(N-trifluoroacetylamino) alkylphosphonic acids

Samples of TFAA-AA^P (1 mmol) were dissolved in 10 ml of appropriate solution (water, 2 M HCl, AcOH/AcONa buffer, 2 M NaOH) containing 10% D₂O and 1 mmol of phosphoric(V) acid or its salts and kept in a thermostat at $20 \pm 0.5\,^{\circ}\text{C}$ (or in the thermoregulated oil bath, in the case of exposition at elevated temperatures). Temporarily ^{31}P NMR spectra of corresponding reaction mixtures were recorded.

Results and discussion

Trifluoroacetylation of 1-aminoalkylphosphonic acids

The acylation reaction of aminoalkylphosphonic acids, despite of the advantage of monitoring by ^{31}P NMR, still belongs to scarcely explored domains of the chemistry of organophosphorus compounds. As a matter of fact, the complicated mechanism of the reactions of AA^P with acylation reagents was the subject of only a few literature reports (Soroka, 1987; Hirschmann et al., 1995, 1997). These revealed competitive course of the acylation, namely N-acylation $[AA^P \rightarrow (AC)-AA^P]$ vs. O-acylation of AA^P , $\{AA^P \rightarrow AA^P[O-(AC)]\}$, occurring as result of the so-called *active sterical hindrance* of the phosphonic function (Scheme 1) (Soroka, 1987).

Tentative investigations on the reaction course of AA^P with anhydrides carried out by us (Kudzin et al., 2005), especially those with an application of the TFAA/TFA reagent (Depczyński, 2006), revealed unexpectedly even far more complex mechanism of these reactions. This is demonstrated in the ³¹P NMR spectra of reaction mixtures of TFAA/TFA with representative 1-aminoalkylphosphonic acids, namely: Ala^P and Pgly^P, presented in Figs. 2 and 3.

$$R' \rightarrow O$$
 $R \rightarrow NH$
 $H \rightarrow P \rightarrow O$
 $H \rightarrow O$

Scheme 1. Reaction course of 1-aminoalkylphosphonic acids with acyl chlorides

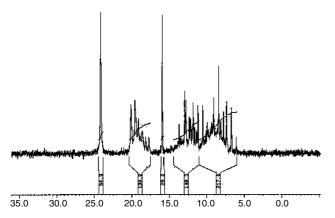


Fig. 2. ^{31}P NMR spectrum of the reaction mixture of Ala^{P} and TFAA/TFA

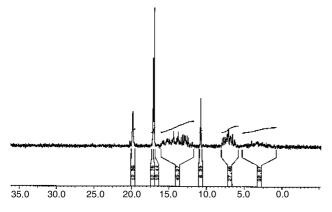


Fig. 3. ^{31}P NMR spectrum of the reaction mixture of Pgly P and TFAA/TFA

Discussed spectra present the multicomponent mixtures containing intermediary compounds 2 – products of N-trifluoroacetylation and O-trifluoroacetylation of AA^P (as singlets), as well as products of their dimerization (singlets and/or dublets) and polycondensation (multiplets) (Scheme 2: $AA^P \rightarrow 2$) (Depczyński, 2006). Corresponding

trimers presumably can exist in both acyclic (2C) and cyclic (2E) forms (Ohms et al., 1992; Diemert et al., 1998).

All investigated reaction mixtures of AA^P with TFAA/TFA converted rapidly into the one ³¹P signal solutions, containing corresponding TFA-AA^P, during their pretreatment with water (Scheme 2: AA^P→2→TFA-AA^P). This enabled a quantitative conversion of 1-aminoalkylphosphonic acids into their TFA-AA^P derivatives, which were isolated in an analytically pure state after direct evaporation of the corresponding post-reaction mixtures. The analytical properties of 1-(*N*-trifluoroacetyl-amino) alkylphosphonic acids obtained are described under Materials and methods.

Deacylation of 1-(N-trifluoroacetylamino) alkylphosphonic acids

Since the trifluoroacetyl function belongs to common amine-protecting groups applied in the peptides chemistry (Weygand and Frauendorfer, 1970; Wunsch, 1994; Kociensky, 1994), we undertook some investigations on its application in the field of phosphonopeptides. In preliminary approach, we started with a search on stability of the trifluoroacetyl-amide function of TFA-AA^P.

Our investigations were performed in 0.1 M aqueous solutions of TFA-AA^P, strongly differing in respect to the pH. These included solutions of TFA-AA^P in water (pH \sim 3.5), in 2 M HCl (pH \sim 0) and 2 M NaOH (pH \sim 14), and also in acetate buffer (pH \sim 4.54) aqueous solutions. All stock solutions were fortified with internal standards consisting of phosphoric acid or appropriate phosphate salts (Na_nH_{3-n}PO₄). The progress of TFA-AA^P deacylations was monitored by ³¹P NMR. Identification of the deacylation products was performed on the basis of chemical shift data of the starting TFA-AA^P and final

Z. H. Kudzin et al.

Table 1. Chemical shifts δ (³¹P) of 1-aminoalkylphosphonic and 1-(*N*-trifluoroacetyl-amino)alkylphosphonic acids

Acids		³¹ P NMR; δ (ppm)			
1 or 3	AA ^P /TFA-AA ^P	2 M HCl ^a	Buffera	H ₂ O ^a	2 M NaOH ^a
1a	Gly ^P	13.9	10.6	11.0	19.3
1b	Ala ^P	16.8	13.9	14.3	22.3
1c	Val ^P	15.7	12.7	13.0	21.2
1d	$Pgly^{P}$	12.5	10.2	10.5	18.0
1e	Phe ^P	14.8	12.1	12.5	20.4
3a	TFA-Gly ^P	18.1	14.1	16.8	
3b	TFA-Ala ^P	21.2	17.5	20.3	
3c	TFA-Val ^P	19.8	16.0	18.9	
3d	TFA-Pgl ^P	16.1	12.9	14.8	
3e	TFA-Phe ^P	18.9	15.5	17.9	
IS	$\begin{array}{c} H_{3}PO_{4}/\\Na_{n}H_{m}PO_{4} \end{array}$	-0.54	+0.12	-0.10	5.39

^a Aqueous solutions of AA^P or TFA- AA^P (0.1 mmol per ml) containing 10% D_2O and phosphoric (V) acid (0.05 mmol per ml) or phosphates (V) (0.05 mmol per ml), applied as internal standards (IS)

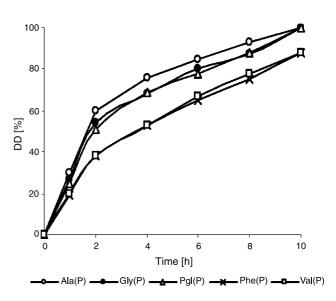
AA^P, determined in appropriate solutions and listed in Table 1.

Deacylation results of representative TFA-AA^P, are illustrated graphically in Figs. 4 and 5.

The graphs represent the plots of corresponding, so the called, deacylation degree (DD) values in function of the reaction time. DD factors, defined by Eq. (1), were directly determined from corresponding ³¹P NMR spectra:

$$DD = \frac{S_{(AAP)}}{S_{(AAP)} + S_{(TFA-AAP)}} \times 100\%$$
 (1)

where: DD – deacylation degree factor; $S_{(AAP)}$ and $S_{(TFA-AAP)}$ – ^{31}P peak areas of starting TFA-AA^P and final



termediary anhydrides 2

Scheme 2. Reaction course of 1-aminoalkylphosphonic acids with trifluoroacetic anhydride with subsequent hydrolysis of in-

Fig. 4. Deacylation course of 1-(*N*-trifluoroacetylamino)alkylphosphonic acids TFA-AA^P (TFA-Gly^P, TFA-Ala^P, TFA-Val^P, TFA-Pgly^P and TFA-Phe^P) in 2 M HCl_{aq} solutions (temp. $100 \pm 0.5\,^{\circ}$ C)

AA^P, determined from ³¹P NMR spectra of corresponding reaction mixtures.

The results obtained exhibited the substantial stability of TFA-AA^P in water and buffer solutions at ambient temperatures (no changes up to 24 h), gradual deacylations in 2 M HCl solutions (Fig. 4) and rapid deacylations in 2 M NaOH solution (100% of deacylation of TFA-AA^P in 15 min). Compounds TFA-AA^P present also the moderate stability in aqueous solutions at elevated temperatures (Fig. 5). These observations can be useful for the protection/deprotection sequences concerning AA^P (AA^P \rightarrow TFA-AA^P \rightarrow AA^P), and applied in the chemistry of aminophosphonates and phosphonopeptides. Moreover,

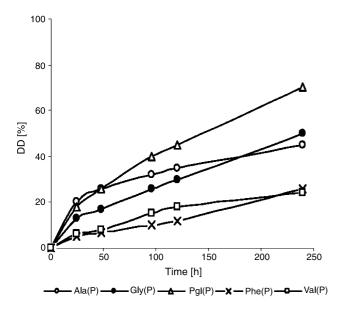


Fig. 5. Deacylation course of 1-(*N*-trifluoroacetylamino)alkylphosphonic acids (TFA-AA^P: TFA-Gly^P, TFA-Ala^P, TFA-Val^P, TFA-Pgly^P and TFA-Phe^P) in aqueous solutions (temp. $20 \pm 0.5\,^{\circ}\text{C}$)

they have to be taken also into consideration during routine work-up procedures of TFA-AA^P (e.g. purifications with active carbon, neutralizations, evaporations of aqueous solutions, etc.).

Conclusion

In summary, the trifluoroacetylation procedure of 1-aminoalkylphosphonic acids we describe, is quantitative, completely general and simple. They make 1-(*N*-trifluoroacetylamino)-alkylphosphonic acids, compounds so far not described (excluding TFA-Val^P), readily available.

The reaction of 1-aminoalkylphosphonic acids with TFAA proved to occur through complex, not described earlier mechanism, postulating the formation of intermediary mixed anhydrides **2**, derived from TFAA and phosphonic, pyrophosphonic and polyphosphonic amino acids $(AA^P \rightarrow 2A-2D)$. These intermediates **2** underwent quantitative conversion to the title 1-(*N*-trifluoroacetylamino)-alkylphosphonic acids during subsequent treatment with water $(2 \rightarrow TFA-AA^P)$. 1-(*N*-Trifluoroacetylamino)alkylphosphonic acids contain labile amide function which can be easily removed during exposition in 2 M NaOH solutions or in boiling water solution $(TFA-AA^P \rightarrow AA^P)$.

A study on antibacterial activities of representative 1-(*N*-trifluoroacetylamino)alkylphosphonic acids is underway.

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